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Antiviral effect of copper chloride on feline calicivirus and synergy with ribavirin in vitro

Dengliang Li^{1†}, Zhanding Cui^{1†}, Guohua Li^{2,3}, Liangting Zhang⁴, Ying Zhang^{2,5}, Han Zhao¹, Shuang Zhang¹, Yanbing Guo⁶, Yanli Zhao¹, Fanxing Men¹, Shihui Zhao¹, Jiang Shao¹, Dongju Du¹, Hailong Huang¹, Kai Wang¹, Guixue Hu^{1*}, Tiansong Li^{7*} and Yongkun Zhao^{2*}

Abstract

Background: Feline calicivirus (FCV) is a common and highly prevalent pathogen causing upper respiratory diseases in kittens and felines in recent years. Due to the substantial genetic variability of the viral genes, existing vaccines cannot provide complete protection. Therefore, research on FCV antiviral drugs has received much attention.

Results: In this study, we found that copper chloride had dose-dependent antiviral effects on FCV in F81 cells. We also found that the combination of copper chloride and ribavirin had a synergistic protective effect against FCV in F81 cells. In contrast, the combination of copper chloride and horse anti-FCV immunoglobulin F (ab')₂ showed an antagonistic effect, likely because copper chloride has an effect on F (ab')₂ immunoglobulin; however, further research is needed to clarify this supposition.

Conclusions: In summary, we found that copper chloride had low cytotoxicity and significant antiviral effects on FCV in F81 cells, providing a new drug candidate for the prevention and treatment of FCV infection.

Keywords: Feline calicivirus, Copper chloride, Antiviral effect, Synergistic protective effect, Antagonistic effect

Background

Feline calicivirus (FCV) is a small, nonenveloped, single positive-strand RNA virus that belongs to the genus *Vesivirus* of the family *Caliciviridae* [1]. Most felines (e.g., cats, tigers, and cheetahs) are susceptible, and infection in dogs has been reported in recent years [2, 3]. Kittens aged 1 to 12 weeks are mainly infected, which

results in a mortality rate of 67% [4, 5]. Infected animals present with oral ulcers, chronic stomatitis, rhinitis, conjunctivitis, and pneumonia [6–9]. The primary prevention method at present is vaccination. However, due to the high evolutionary rate of the FCV capsid protein, which results in as many as 1.3×10^{-2} to 2.6×10^{-2} substitutions per nucleotide per year [10], traditional vaccination cannot completely protect animals from reinfection with mutant or wild-type strains, even though FCV is considered to have only one serotype [11]. Therefore, it is necessary to develop a safe and effective antiviral drug as a monotherapy or as part of a combination treatment.

In previous studies, various compounds had been found to have anti-FCV effects, such as minomorpho

* Correspondence: huguixue901103@163.com; its0309@163.com; zhaoyongkun1976@126.com

[†]Dengliang Li and Zhanding Cui are joint first authors to this paper.

¹College of Animal Science and Technology, Jilin Agricultural University, Changchun 130118, China

²College of Science, Beihua University, Jilin 132013, China

³Institute of Military Veterinary Medicine, Academy of Military Medical Sciences, 666 Liuying West Road, Changchun 130122, China

Full list of author information is available at the end of the article



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oligophosphate (PMO), lithium chloride, and germanone [12–14]. The combination of mefloquine and recombinant cat interferon- ω had a synergetic effect against FCV [15]. Copper is an indispensable element used in the production of livestock and poultry. Basic copper chloride is an essential additive in feed [16]. Researchers previously found that copper and copper compounds had antiviral effects against dengue virus, influenza virus, and human immunodeficiency virus (HIV) in vitro [17–19]. A research report on the cytotoxicity of copper chloride showed that the cytotoxicity of copper chloride was minimal in the range of cytotoxic concentrations that did not disrupt the stability of copper [20]. The copper (II) chloride complex has also been utilized for its anticancer effects in vitro on human cervical cancer, colon cancer, ovarian cancer, and melanoma cell lines [21]. However, there have been no reports of the antiviral effect of copper chloride against FCV.

In this study, we found that copper chloride had low cytotoxicity in F81 cells. We evaluated the antiviral effects of copper chloride in vitro in a dose-dependent manner by measuring the TCID₅₀ and using RT-qPCR to analyse the effects against FCV. Additionally, the combination of copper chloride and ribavirin showed synergistic antiviral effects against FCV.

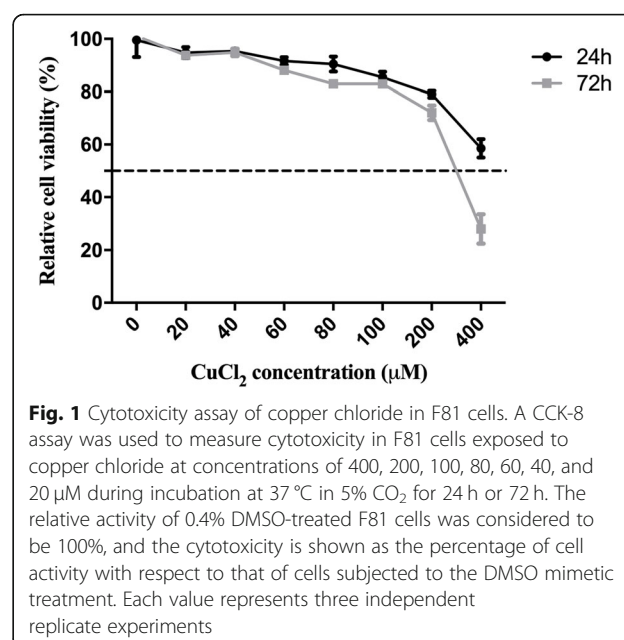
Results

Cytotoxicity test of copper chloride in F81 cells

The results of the cytotoxicity assay showed that the relative cell viability was greater than 85% after treatment with copper chloride at a concentration of 20 ~ 80 μ M for 24 h or 72 h. When the concentration of copper chloride was less than or equal to 200 μ M (below the CC₅₀), the relative cell viability of the F81 cells was higher than 50% after 24 h or 72 h of treatment, indicating that copper chloride can be regarded as nontoxic in this concentration range (Fig. 1). Therefore, a concentration of 200 μ M copper chloride was used as the maximum concentration for the antiviral experiments.

Antiviral effect of copper chloride at different doses

To evaluate the antiviral effects of different doses of copper chloride on FCV, we examined the virus titer and RNA levels of FCV in F81 cells treated with different doses of copper chloride. The results showed that the virus titre was significantly lower than that of the mock group in cells exposed to 60, 80, 100, and 200 μ M concentrations of copper chloride ($p < 0.01$) (Fig. 2a). The relative RNA levels of FCV were significantly reduced compared to those of the mock group when the copper chloride concentration was 80 or 100 μ M ($p < 0.05$), and the decrease was highly significant at 200 μ M copper chloride ($p < 0.001$) (Fig. 2b). In addition, all the results indicated that the antiviral effect of copper chloride on



F81 cells was dose-dependent. The IC₅₀ of copper chloride for FCV was determined to be 5.1 μ M (Fig. 2c).

Indirect immunofluorescence assay

To further evaluate whether the antiviral effect of copper chloride on FCV was dose-dependent, we performed an indirect immunofluorescence assay (IFA). The results showed that an intense green fluorescence signal was observed in the 0 μ M group, and only weak fluorescence signals were observed at concentrations of 60, 40, and 20 μ M, indicating a dose-dependent relationship after greyscale scanning (Fig. 3b). There was almost no fluorescence signal at concentrations of 80, 100, and 200 μ M or for the control group compared to that of the mock group (Fig. 3a). In addition, the F81 cells treated with 200 μ M copper chloride showed slight rounding and polymerization. This finding also validates the previously described range of the nontoxic concentrations of copper chloride.

Antiviral effect of copper chloride for different treatment times

To evaluate the antiviral effect of copper chloride on FCV for different treatment times, we examined the virus titer and RNA levels of FCV in F81 cells for different treatment times. The experimental data showed that 80 μ M copper chloride had good antiviral effects on FCV and decreased cytotoxicity. Therefore, we selected this concentration of copper chloride for subsequent experiments. The results showed that the virus titre and the relative RNA level of FCV were significantly lower in the copper chloride treatment group after -1, 0, and 1 h of FCV infection than those in the mock group ($p < 0.001$), and there was also a significant difference after 4

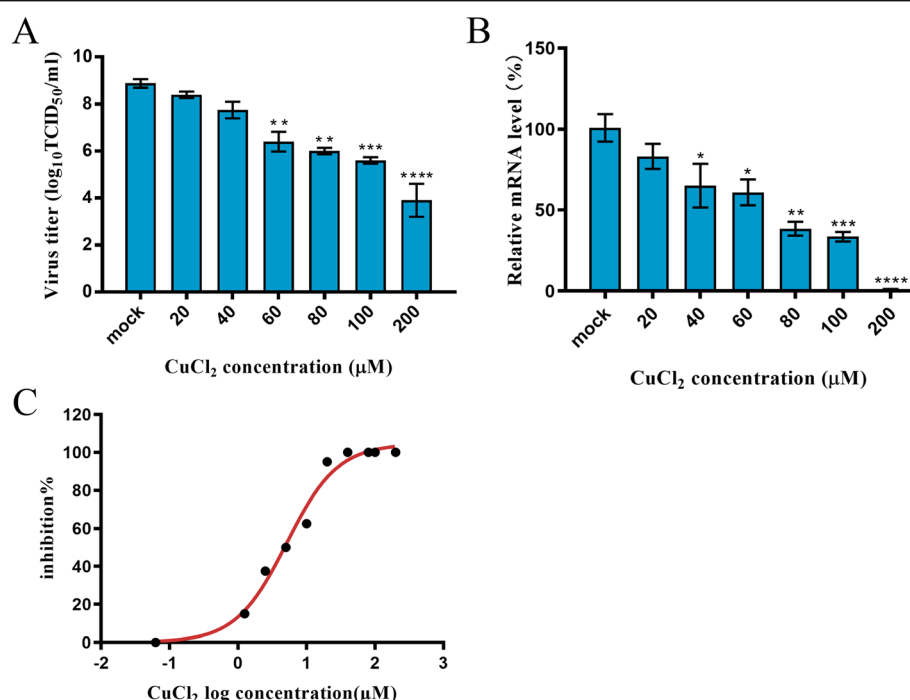


Fig. 2 The antiviral effect of different doses of copper chloride (20–200 μM) on F81 cells infected with FCV at 100 TCID₅₀. After incubation for 28 h at 37 °C in 5% CO₂, the virus titre (a) and the relative RNA levels (b) of FCV were detected. c The IC₅₀ of copper chloride for FCV was determined; * $p < 0.0332$; ** $p < 0.0021$; *** $p < 0.0002$; and **** $p < 0.0001$

and 8 h of infection ($p < 0.002$). After 16 h of virus infection, there were no significant differences between the virus titre or relative RNA level of the FCV and the mock group, indicating that the antiviral effect of copper chloride (80 μM) was not significant after 16 h of FCV infection (Fig. 4a and b). The RT-qPCR data also reflected similar results. Thus, the antiviral effect of copper chloride on FCV was mainly observed in the early stages of virus proliferation.

Antiviral effect of copper chloride on different strains of FCV

For other strains of FCV, the antiviral effects of copper chloride were validated. As previously described, we determined the viral titre and relative viral RNA levels of the different strains after copper chloride treatment. The results showed that the use of 80 μM copper chloride to treat other strains of FCV (CH-JL1, CH-JL3, CH-JL4, and CH-SH) significantly reduced the viral titre and relative viral RNA levels of the infected F81 cells (Fig. 5a and b). These results indicated that copper chloride had a strong antiviral effect against strains of FCV with differing genotypes.

Synergistic effect of ribavirin and the antagonistic effect of F (ab')₂

The checkerboard method was used to mix different compounds, and the antiviral effect of the compound

combinations were evaluated using SynergyFinder. The results showed that the combination of copper chloride and ribavirin had a synergistic protective effect on FCV-infected F81 cells within a specific concentration range (Fig. 6a). The ZIP model was used to calculate the average score of the synergy of the two compounds, and it was found to be 9.687 (Fig. 6b). Conversely, the combined use of copper chloride and F (ab')₂ showed antagonistic effects, and the average score of the two compounds calculated by the ZIP model was −20.798 (Fig. 6c and d).

Discussion

In previous studies, copper or copper ions were shown to have an antiviral effect on herpes simplex virus (HSV) [22]. However, they were mainly used to inactivate viruses since copper ions can disrupt the activity of certain proteins containing hydroxyl radicals [21, 23]. However, when the concentration of copper ions was too high, the resulting toxicity was fatal because copper chloride can inactivate specific proteins and produce a large number of reactive oxygen species in cells to induce apoptosis [24]. In our study, we found that low concentrations of copper chloride induced low levels of cytotoxicity in F81 cells. The F81 cells treated with copper chloride concentrations up to 200 μM for 24 h and 72 h showed cell activity greater than 50%, which was similar to that

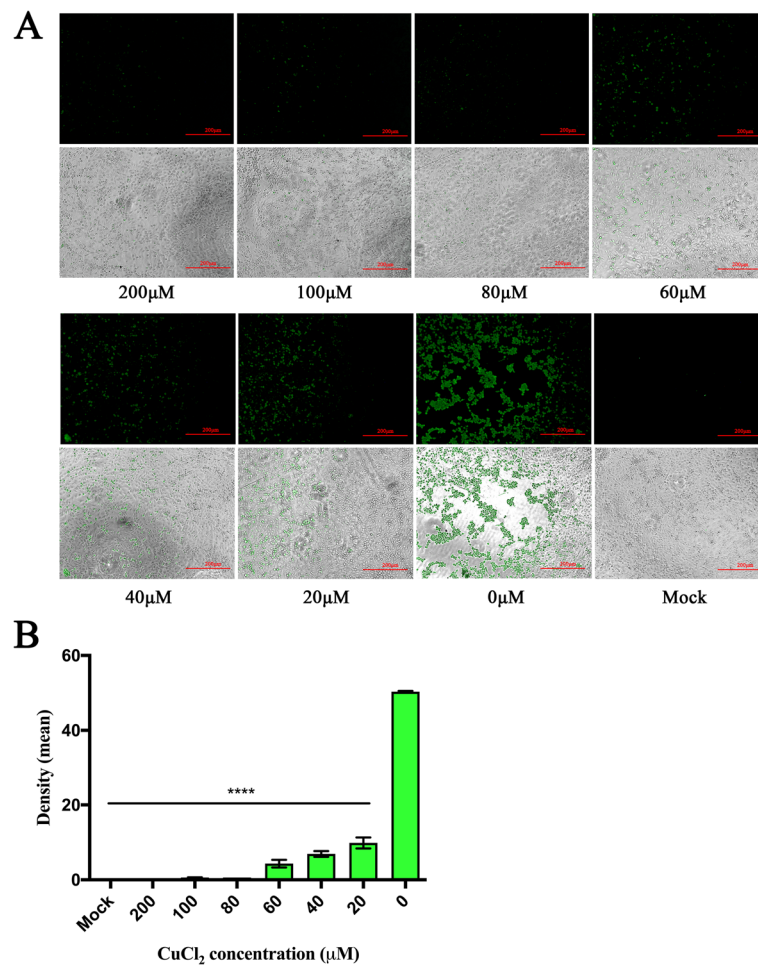


Fig. 3 An IFA verified the antiviral effect of copper chloride. **a** F81 cells were infected with different concentrations of copper chloride (20–200 μM) and FCV (100 TCID₅₀), and a mock treatment group containing 0.4% DMSO and a negative control group that was not infected with FCV were used as controls. After incubation for 1 h at 37 °C in 5% CO₂, an IFA of the F81 cells was performed. **b** Two experimental wells were selected in the 96-well cell plate, and the fluorescence signals were detected in randomly selected areas in each well using ImageJ software. The experimental result was expressed as the average of two fluorescence signals. The antiviral effect of copper chloride was evaluated by detecting the fluorescence signal; **** $p < 0.0001$

previously reported for the cytotoxicity of soluble copper chloride. Therefore, the cytotoxicity of copper chloride was very low within the range of cytotoxic concentrations that did not disrupt the stability of copper [20]. However, the safety of copper chloride administration to animals remained to be further explored. Previously, copper chloride dihydrate treatment exhibited excellent antiviral activity against dengue virus, but the antiviral effect of copper chloride on FCV had not been reported [25]. In this study, we found that copper chloride had an antiviral effect on FCV with an IC₅₀ of 5.1 μM. The IFA results showed that the inhibition of FCV by copper chloride was dose-dependent and significantly reduced the cytopathic effect (CPE) of FCV on F81 cells. After treatment with different concentrations of copper chloride, the viral titer and RNA expression levels of FCV in

cells were significantly different from those in the control cells.

Then, we demonstrated the inhibitory effect of copper chloride on the viral titre and RNA levels of FCV at different time points. FCV replication was significantly inhibited when copper chloride was added 8 h after FCV was used to infect cells, but copper chloride did not significantly inhibit FCV after 16 h. Our findings were similar to those of previous studies using fexaramine, LiCl, ginger extracts, or CSX [13, 14, 26, 27]. We speculated that copper chloride could hinder the attachment and entry of FCV and inhibit replication inside the cell, but the mechanism involved in inhibiting replication needs further verification. Past research has shown that copper ions have the ability to disrupt DNA through cross-linking with single-stranded polynucleotide chains as

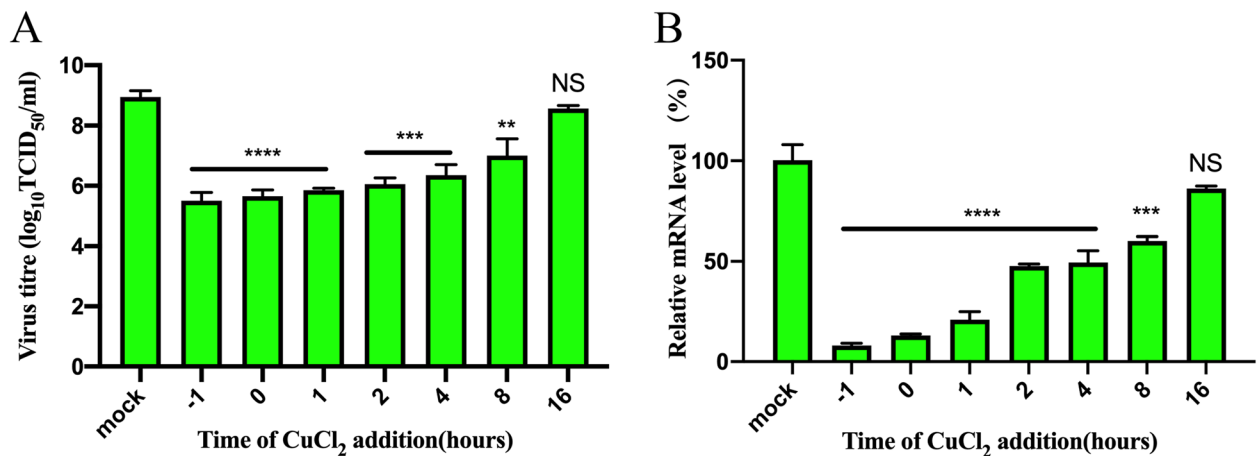


Fig. 4 The antiviral effect of copper chloride on FCV at different time points. F81 cells were infected with 100 TCID₅₀ FCV and then treated with 80 μ M copper chloride for -1, 0, 1, 2, 4, 8, and 16 h. After incubation for 24 h at 37 °C in 5% CO₂, the virus titre (a) and relative RNA levels (b) of FCV were detected. NS, $p > 0.1234$; ** $p < 0.0021$; *** $p < 0.0002$; and **** $p < 0.0001$. ('-1 h' indicates that the cells were treated with CuCl₂ 1 h before infection)

well as forming intra- and intra-strand cross-links [26]. Numerous studies have reported that copper ions can inhibit neuraminidase (NA) in cells and tissues [27, 28]. Both influenza virus NA [17, 29] and Sendai virus NA [30] could be inhibited by copper, but it had not been determined whether the antiviral effect was related to NA inhibition. The copper ions resulted in the abnormal morphology of the virus, which might indicate that the structural changes could be related to antiviral effects [17]. We found that copper chloride also inhibited the replication of other FCV strains in F81 cells, which was further evidence of the antiviral activity of copper chloride against FCV. The ZIP model in SynergyFinder was used to analyse the combined effects of the drugs. The

synergy score for the ZIP model was expressed as the average of all the δ scores in the dose-response landscape [25, 31]. In a later experiment, we found that the broad-spectrum antiviral drug ribavirin and copper chloride had synergistic effects on inhibiting FCV. The most synergistic area had a score of 15.5, with an average score of 9.7. However, copper chloride and immunoglobulin F (ab')₂ showed a robust antagonistic effect. It has been reported in the literature that copper ions could induce IgG cleavage [32, 33]. Past research has found that when metal ions with a specific stereochemistry in the appropriate conformation are present, metal ions can increase the rate of peptide cleavage and catalyse the reaction [34, 35]. In fact, copper ions could bind

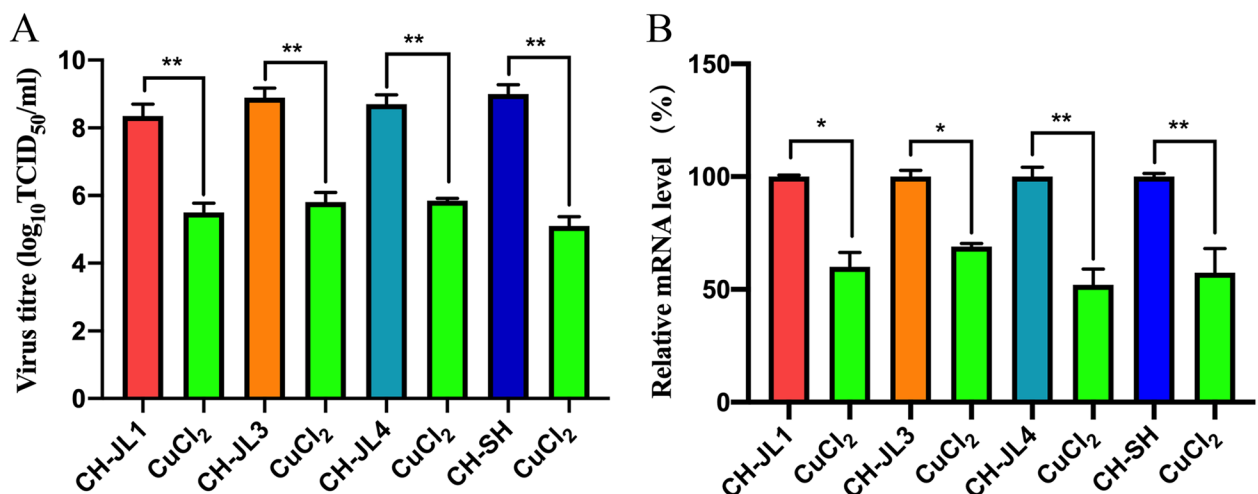
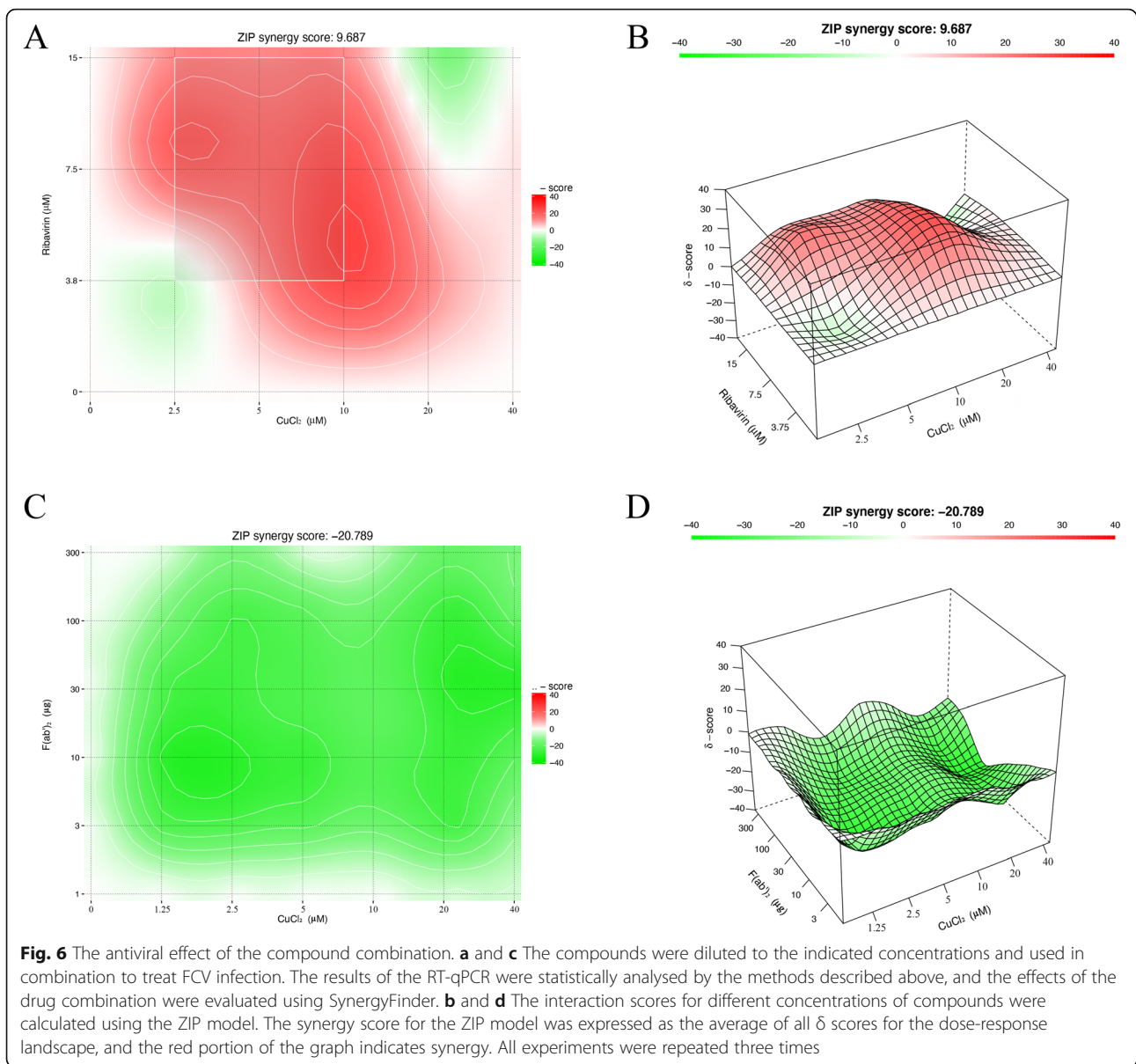


Fig. 5 The antiviral effect of copper chloride on different FCV strains. The F81 cells were infected with different FCV strains at 100 TCID₅₀ and treated with 80 μ M copper chloride. After incubation for 28 h at 37 °C in 5% CO₂, the virus titre (a) and the relative RNA levels (b) of FCV were detected; * $p < 0.1234$ and ** $p < 0.0021$



and enhance the degradation of proteins such as monoclonal antibodies and hydrolyse small peptides and bovine serum albumin, and trace amounts of copper ions might be sufficient to enable site-specific, metal-mediated monoclonal antibody degradation [36]. When copper chloride and immunoglobulin F (ab')₂ are applied to F81 cells infected with FCV at the same time, there might be mutual effects that cause antagonistic activity. However, further research is required to prove this hypothesis.

Conclusions

In summary, we verified that copper chloride had low cytotoxicity in F81 cells and had an antiviral effect on FCV in vitro. Later, we found that copper chloride had

an effect on the replication stage in FCV-infected cells, and it had a synergistic effect with ribavirin, but drug resistance should be determined in a future study. This study indicated that copper chloride could be a promising candidate drug for the treatment of FCV infection in felines.

Methods

Viruses, cells and compounds

The FCV CH-SH strains and F81 cells were provided by the Institute of Military Veterinary Medicine, Academy of Military Medical Science. FCV CH-JL1/CH-JL2/CH-JL3/CH-JL4 were isolated and stored in our laboratory [37]. The horse anti-FCV immunoglobulin F (ab')₂ was produced and stored in our laboratory [38]. The copper

chloride dihydrate (Aladdin, China) product number was C111685. The ribavirin (Aladdin, China) product number was R101754.

Cytotoxicity assay for copper chloride

Copper chloride was diluted in DMSO to a concentration of 100 mM as a stock solution, and the stock solution was diluted to different concentrations (400 μ M, 200 μ M, 100 μ M, 80 μ M, 60 μ M, 40 μ M, and 20 μ M) with MEM containing 2% FBS. The cells were incubated at 37 °C in 5% CO₂ for 24 h or 72 h. After washing the cells twice with PBS solution, 180 μ L of FBS-free MEM and 20 μ L of CCK-8 reagent (CCK-8; Dojindo, Japan) were added to each well. After incubation at 37 °C for 1–2 h, the optical density (OD) was determined at 450 nm using a Cmax Plus microplate reader (Molecular Devices, USA). The cell viability was calculated by the formula $[\text{OD}_{450}(\text{compound}) - \text{OD}_{450}(\text{blank})] / [\text{OD}_{450}(\text{control}) - \text{OD}_{450}(\text{blank})] \times 100\%$. The copper chloride concentrations that were less than 50% of the cytostatic concentration (CC₅₀) were defined as nontoxic [39].

Virus titre and genome detection

The virus solution to be detected was diluted serially 10-fold. Each concentration solution was added to each column at 100 μ L per well, which was repeated 8 times for each concentration, followed by the addition of 100 μ L of 2% MEM. The control was established in virus-free medium and cultured at 37 °C in 5% CO₂. The virus TCID₅₀ was calculated using the Reed and Muench formula. Relative RT-qPCR was used to evaluate FCV gene expression. Briefly, first, the RNA of the virus was extracted from the mixture of cells and medium (by freezing and thawing three times) with the Simply P Total RNA Extraction Kit (Bioflux, China); then, the RNA was reverse-transcribed into complementary DNA (cDNA) (Thermo, USA), followed by RT-qPCR using TB Green Premix Ex Taq II (Tli RNaseH Plus) (TaKaRa, China). The RT-qPCR conditions were based on the Applied Biosystems 7500 Fast Real-Time PCR System with TB Green Premix Ex Taq II (Tli RNaseH Plus) (TaKaRa, China). The upstream and downstream sequences of the FCV primers were 5'-GCAGGTTGGGATAAACATGGA-3' and 5'-CACGAGGCGATTGAGTTGAG-3', and for GAPDH, the sequences were 5'-TGGAAAGCCCATCACCATC-3' and 5'-ACTCCACAACATAC TCAGCACCA-3'.

Antiviral effect of copper chloride on FCV

F81 cells at 100% confluence were infected with FCV CH-JL2 at 100 TCID₅₀, while 200 μ L of different concentrations of copper chloride (200–20 μ M) were added to the medium. MEM containing 0.4% DMSO was used for the mock treatment group. After incubation for 28 h

at 37 °C in 5% CO₂, the TCID₅₀ levels of the virus and the RT-qPCR results were determined to assess the antiviral effect on FCV. The half-maximal inhibitory concentration (IC₅₀) of copper chloride for FCV was also determined, and the results were plotted using Graph-Pad Prism 8.

Indirect immunofluorescence assay (IFA)

To more intuitively observe the antiviral effect of copper chloride on FCV, we performed an IFA [38]. As described above, a monolayer of F81 cells infected with 100 TCID₅₀ of FCV was exposed to different concentrations of copper chloride and then fixed in 80% cold acetone for 30 min. Next, the cells were washed 5 times with PBS containing 0.05% Tween-20 (PBST). Then, primary antibody (VMRD, USA) diluted 300-fold with 1% BSA was added, and the cells were incubated for 1 h at 37 °C. The supernatant was discarded, the cells were washed 5 times with PBST, and 200-fold diluted FITC-labelled rabbit anti-cat secondary antibody (Bioss Antibody, China) was added to the cells in the dark and incubated at 37 °C for 1 h. The fluorescence was observed under an inverted Leica DMI8 fluorescence microscope (Leica, Germany), and the fluorescence results were analysed by greyscale scanning.

Effect of different copper chloride treatment times on FCV

To determine whether the antiviral effect of copper chloride on F81 cells was time-dependent, copper chloride solution at a final concentration of 80 μ M was administered –1 h, 0 h, 2 h, 4 h, 6 h, 8 h, and 16 h after FCV infection. MEM containing 0.4% DMSO was used for the mock treatment group. After incubation for 24 h at 37 °C in 5% CO₂, the TCID₅₀ level was determined, and the RT-qPCR results were assessed to determine the amount of virus.

Antiviral effect of copper chloride on different strains

To determine whether copper chloride also has antiviral effects on other FCV strains, we diluted different strains of FCV (CH-JL1, CH-JL3, CH-JL4, and CH-SH) to 100 TCID₅₀ and infected F81 cells, which were subsequently treated with copper chloride solution at a final concentration of 80 μ M. After incubating the cells with virus for 28 h at 37 °C in 5% CO₂, the TCID₅₀ level was determined, and the RT-qPCR results were assessed to determine the amount of virus.

Combination of copper chloride and ribavirin or F(ab')₂

To evaluate the combined action of the compounds, we diluted the different drugs by the checkerboard method and mixed the two drugs thoroughly to determine the combined effect on FCV of different concentrations of

copper chloride (0–40 μM) and ribavirin (0–15 μM)/F(ab')₂ (0–300 μg). The results from the RT-qPCR experiments were statistically analysed in a manner described previously, and the effects of the drug combinations were evaluated using SynergyFinder [31]. The zero-interaction efficiency (ZIP) model [25] was used to calculate the mutual scores of the different concentrations of the drug. For each treatment, at least three data sets from triplicate independent experiments were analysed. The results are expressed as the mean and standard error of the mean (SEM).

Statistical methods

All experiments were performed three times independently in triplicate. The data are expressed as the mean \pm standard deviation (SD). The significance of the differences between the groups were determined by paired t-tests and one-way/two-way analysis of variance.

Abbreviations

PMO: Minomorpho oligophosphate; TCID₅₀: 50% tissue culture infective dose; RT-qPCR: Real-time quantitative polymerase chain reaction; CC₅₀: 50% cytostatic concentration; PBS: Phosphate-buffered saline; IC₅₀: Half-maximal inhibitory concentration; IFA: Indirect immunofluorescence assay; SEM: Standard error of the mean; FITC: Fluorescein isothiocyanate; ZIP: Zero-interaction efficiency; F81 cells: Feline kidney fibroblast (F81) cells

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Authors' contributions

LDL, CZD drafted the manuscript and assisted with the conception and design of the study, data collection, and data analysis. LGH, ZLT, ZY, ZH assisted with study design and protocol development. ZS, GYB, ZYL, MFX, ZSH assisted with data analysis. SJ, DDJ, HHL, and WK assisted with interpretation of the study and assisted in writing and editing the manuscript. HGX, LTS and ZYK delineated the hypothesis, helped conceive and design the study, performed and oversaw the data analyses, and assisted in the writing of the manuscript. All authors read and approved the final manuscript for submission and publication.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author upon reasonable request.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹College of Animal Science and Technology, Jilin Agricultural University, Changchun 130118, China. ²Institute of Military Veterinary Medicine, Academy of Military Medical Sciences, 666 Liuying West Road, Changchun 130122, China. ³College of Animal Science and Technology, Shihezi

University, Shihezi 832003, China. ⁴College of Continuing Education, Hebei Agricultural University, Baoding 071001, China. ⁵College of Wildlife and Protected Area Northeast Forestry University, Harbin 150040, China. ⁶Animal Husbandry and Veterinary Science Research Institute of Jilin Province, Changchun 130062, China. ⁷College of Science, Beihua University, Jilin 132013, China.

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