

METHODOLOGY ARTICLE

Open Access

Use of a Western blot technique for the serodiagnosis of glanders

Mandy C Elschner^{1*}, Holger C Scholz², Falk Melzer¹, Muhammad Saqib^{3,4}, Peggy Marten¹, Astrid Rassbach¹, Michael Dietzsch¹, Gernot Schmoock¹, Vania L de Assis Santana⁵, Marcilia MA de Souza⁵, Renate Wernery⁶, Ulrich Wernery⁶, Heinrich Neubauer¹

Abstract

Background: The in vivo diagnosis of glanders relies on the highly sensitive complement fixation test (CFT). Frequently observed false positive results are troublesome for veterinary authorities and cause financial losses to animal owners. Consequently, there is an urgent need to develop a test with high specificity. Hence, a Western blot assay making use of a partly purified lipopolysaccharide (LPS) containing antigen of three *Burkholderia mallei* strains was developed. The test was validated investigating a comprehensive set of positive and negative sera obtained from horses and mules from endemic and non endemic areas.

Results: The developed Western blot assay showed a markedly higher diagnostic specificity when compared to the prescribed CFT and therefore can be used as a confirmatory test. However, the CFT remains the test of choice for routine testing of glanders due to its high sensitivity, its feasibility using standard laboratory equipment and its worldwide distribution in diagnostic laboratories.

Conclusions: The CFT should be amended by the newly validated Western blot to increase the positive likelihood ratio of glanders serodiagnosis in non endemic areas or areas with low glanders prevalence. Its use for international trade of horses and mules should be implemented by the OIE.

Background

Glanders, caused by *Burkholderia (B.) mallei*, is a highly contagious disease in equines which is notifiable to the World Organisation of Animal Health (OIE, Office International des Epizooties). The disease is still endemic in the Middle East, Asia and South America. Recent outbreaks have been reported from Turkey, the United Arab Emirates, Iraq, Iran, India, Pakistan, Mongolia, China, Brazil and most recently from Bahrain [1-11]. The distribution of glanders in Africa is unknown.

The diagnosis of *B. mallei* infection still relies on serological proof by agglutination test and complement fixation test (CFT), or proof of the presence of a specific delayed hypersensitivity reaction after intracutaneous application of mallein [12,13]. The CFT for glanders is so far the only officially recognized serological test in

international trade of equidae. The CFT has a sensitivity of at least 97% [14] but a notable number of unspecific, false positive results occur [15-18]. False positive results due to cross-reactions may be seen in horses suffering from strangles, equine influenza or petechial fever. The test can also not be applied on sera having so called "anticomplementary activity". In general, serological tests may be negative in emaciated and chronically debilitated animals suffering from glanders [16].

Glanders was eradicated from Western Europe, Australia and North America in the last century applying a rigorous culling of horses found positive in complement fixation and mallein test [15,19]. These techniques are useful in eradication programs with regard to specificity and sensitivity [12]. However, in glanders free areas or in areas with very low prevalence of glanders highly specific tests are needed to minimize the number of false positive results [20]. Lipopolysaccharide (LPS) preparations have already been applied in Western blot analysis or competition ELISA to detect anti-*B. mallei* antibodies in serum of horses [20-22]. But current protocols for

* Correspondence: mandy.elschner@fli.bund.de

¹Friedrich Loeffler Institute, Federal Research Institute for Animal Health, Institute of Bacterial Infections and Zoonoses, Naumburger Strasse 96a, 07743 Jena, Germany

Full list of author information is available at the end of the article

the extraction and purification of LPS from *B. mallei* are time consuming, sophisticated and hazardous for the operator [23,24].

Here we describe a new Western blot assay based on an easy to prepare LPS preparation containing antigen from *B. mallei*, purified from soluble components. With a specificity and a sensitivity of 100%, the assay can be used to confirm positive and at the same time to exclude false positive CFT results.

Methods

Production of hyperimmune sera in rabbits

One rabbit was immunized with a crude suspension (10^6 cfu/ml) of heat inactivated *B. mallei*, *B. pseudomallei*, *B. cepacia*, and *Pseudomonas aeruginosa*, respectively. The strains used are given in Table 1. The first and second immunizations (days 0 and 3) were done by intracutaneous application of 10 doses of 0.2 ml (10^6 cfu/ml) antigen without adjuvant distributed on the back. Further, 5 immunizations were done by application of 0.5 ml (10^6 cfu/ml) in the ear vein (days 10, 17, 24, 31, and 38). Seroconversion was monitored by CFT and Western blot assay using the homologous antigen. The final sera were collected 6 weeks after the first immunization (day 42). The animal experiment was authorized by the government of Thuringia, Germany (registration number 04-106/07).

Immunisation of a horse

To produce a panel of positive control sera, one horse was immunized subcutaneously using a mixture of crude suspensions of heat inactivated *B. mallei* strains Mukteswar, Bogor and Zagreb (10^9 cfu/ml) adjuvanted with aluminium hydroxide gel (Sigma Chemie GmbH, Munich, Germany). Immunizations were performed weekly for seven weeks with 7 doses of 1.5 ml antigen containing increasing cell concentrations (10^3 cfu/ml to 10^9 cfu/ml). The titer was analysed in parallel by CFT and Western blot analysis twice a week and for 10 weeks after the first immunization. The animal experiment was authorized by the government of Thuringia, Germany (registration number 04-105/07).

Table 1 List of bacterial strains used in this study

Strain	Source	Geographic origin	Species
Mukteswar	Horse	India	<i>Burkholderia mallei</i>
Bogor	Horse	Indonesia	<i>Burkholderia mallei</i>
Zagreb	Horse	Yugoslavia	<i>Burkholderia mallei</i>
ATCC 23343	Human	unknown	<i>Burkholderia pseudomallei</i>
DSM 7288	unknown	unknown	<i>Burkholderia cepacia</i>
ATCC 9027	unknown	unknown	<i>Pseudomonas aeruginosa</i>

ATCC: American Type Culture Collection, Manassas, USA.

DSM: German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany.

Samples

A total of 2,282 sera (group I) were collected from different horse populations in several geographical areas of Germany during 2006 to 2009.

These sera were considered negative for glanders because Germany is free of the disease for more than 50 years.

The specificity testing was carried out with 305 samples (group II) randomly selected from group I regarding the Mersenne Twister method by use of the Software SPSS (SPSS Inc., Chicago, USA).

The sensitivity of the Western blot assay was tested using 205 true positive sera (group III). Of these, 171 CFT positive samples were collected from microbiologically or clinically/mallein positive glanderous horses and mules in Punjab, Pakistan (n = 59), Pernambuco, Brazil (n = 87), and Dubai, United Arab Emirates (n = 25). Included in group III were additionally 21 and 13 sera from the *B. mallei*-immunized horse and rabbit, respectively.

CFT

The CFT was performed according to the instructions of the OIE Manual of Diagnostic tests and Vaccines for Terrestrial Animals [13] using a certified, commercially available antigen containing antigen from *B. mallei* strains Bogor, Mukteswar, and Zagreb (cc-pro GmbH, Oberdorla, Germany), and a complement and ready-to use hemolytic system (Institut Virion/Serion GmbH, Würzburg, Germany). Samples were considered negative when 100% haemolysis occurred at 1:5 dilution, suspicious when 25-75% haemolysis was seen at dilution 1:5 and positive when no haemolysis was detected at dilution 1:5.

Antigen preparation for the Western blot assay

B. mallei strains Bogor, Zagreb and Mukteswar were grown on blood agar plates over night at 37°C under aerobic conditions. Details on the strains are given in Table 1. For LPS purification, a 10-microliter loop full of bacterial colony material was re-suspended thoroughly in 6 ml saline (0.9% NaCl, pH 7.0) up to a density comparable to McFarland scale 4.0. A volume of 3 ml of 37% formaldehyde was added to achieve a final formaldehyde concentration of 12.3%. The suspension was rigorously vortexed and subsequently swayed using an orbital shaker (Polymax 1040, Heidolph Instruments GmbH, Schwabach, Germany) over night at room temperature. Controls for sterility confirmed inactivation of bacteria. Cells were pelletized by centrifugation at $3.500 \times g$ for 15 min and the supernatant was discarded. Soluble components were removed by 3 consecutive centrifugation and washing steps with 9 ml phosphate buffered saline (PBS, pH 7.0) shaking at 15 rpm for 10 minutes. The LPS-containing pellet was re-suspended in

9 ml PBS. Two further extraction steps by adding 1 volume of 37% formaldehyde into 2 volume re-suspended cells in PBS were applied. By 3 consecutive washing steps with PBS the soluble cell components and the formaldehyde were removed. After the last washing step the pellet was resuspended in PBS. Aliquots were directly used for SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and subsequent Western blot analysis. The washing steps and the number of bacteria used for LPS preparation were crucial for the efficient removal of soluble components from the LPS-containing cell debris. Optimal purification was achieved using 9×10^8 cells/ml (McFarland scale 3) or less (data not shown).

The correct formation of a typical LPS ladder was proofed by SDS-PAGE and silver staining before blotting. For SDS-PAGE several dilutions from the prepared LPS containing cell suspension in Laemmli Buffer (Sigma, Munich, Germany) were loaded on a precast denaturing 4-12% polyacrylamide gradient gel (Invitrogen, Karlsruhe, Germany) and separated at a constant voltage of 200V for 50 min using the Xcell SureLock™ MiniCell (Invitrogen). Silver staining was carried out with the SilverQuest™ silver staining kit (Invitrogen). The LPS extracted from each of the *B. mallei* strains showed the typical LPS ladder with no visible differences. We believe that soluble antigens are mainly removed from the preparation but we are well aware, that our antigen is not pure and contains other components, like high molecular weight components (Figure 1). The protein content of each antigen preparation was below 0.5 mg/ml, measured by absorption at 280 nm (Spectrophotometer NanoDrop 1000, Thermo Fisher Scientific, Bonn, Germany). For further use, the antigen preparations of the *B. mallei* strains Bogor, Zagreb and Mukteswar were mixed equally.

Immuno blotting

For Western blotting analysis, 350 µl LPS-containing suspension of *B. mallei* strains Bogor, Zagreb and Mukteswar were separated on a precast preparative 4-12% polyacrylamide gradient gel (Invitrogen) as described above. LPS was transferred to a 0.45 µm nitrocellulose membrane (Invitrogen) by blotting at constant 30V for 1 h using the Novex® Blot Module (Invitrogen). The membrane was blocked overnight in Blocking Solution (Candor Bioscience, Weißensberg, Germany). After three washing steps the membrane was cut into strips of 3 mm and stored at -20° C or was used for subsequent immunoblot analysis. Stripes were incubated with equine or rabbit sera in a 1:50 dilution in Low Cross Buffer (Candor Bioscience) for 1.5 h at room temperature followed by three washing steps in washing buffer (Candor Bioscience) 20 min each. The strips were then incubated for 1.5 h at room temperature in Low Cross Buffer containing alkaline phosphatase-

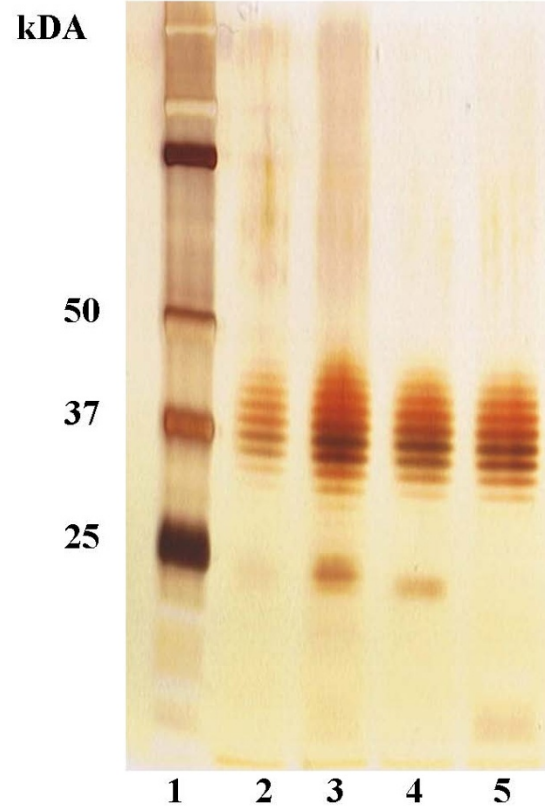


Figure 1 Silver stained SDS-PAGE gel. Silver stain of SDS-PAGE of mixtures of purified LPS of *B. mallei* strains protein standard (lane 1), Mukteswar (lane 2); Bogor (lane 3); Zagreb (lane 4), mixture of all (lane 5).

conjugated rabbit anti-horse-IgG or goat anti-rabbit-IgG (Sigma, Munich, Germany, 1:5,000). After three additional washing steps, the strips were stained with NBT-BCIP® solution (Sigma). The immunostaining was stopped by washing with distilled water after 10 minutes. The immunoblot was scored positive if the banding pattern of the *B. mallei* LPS ladder within the region of 20 to 60 kDa was clearly visible, scored suspicious if a weak color reaction was detected, and scored negative if no reaction was seen (Figure 2).

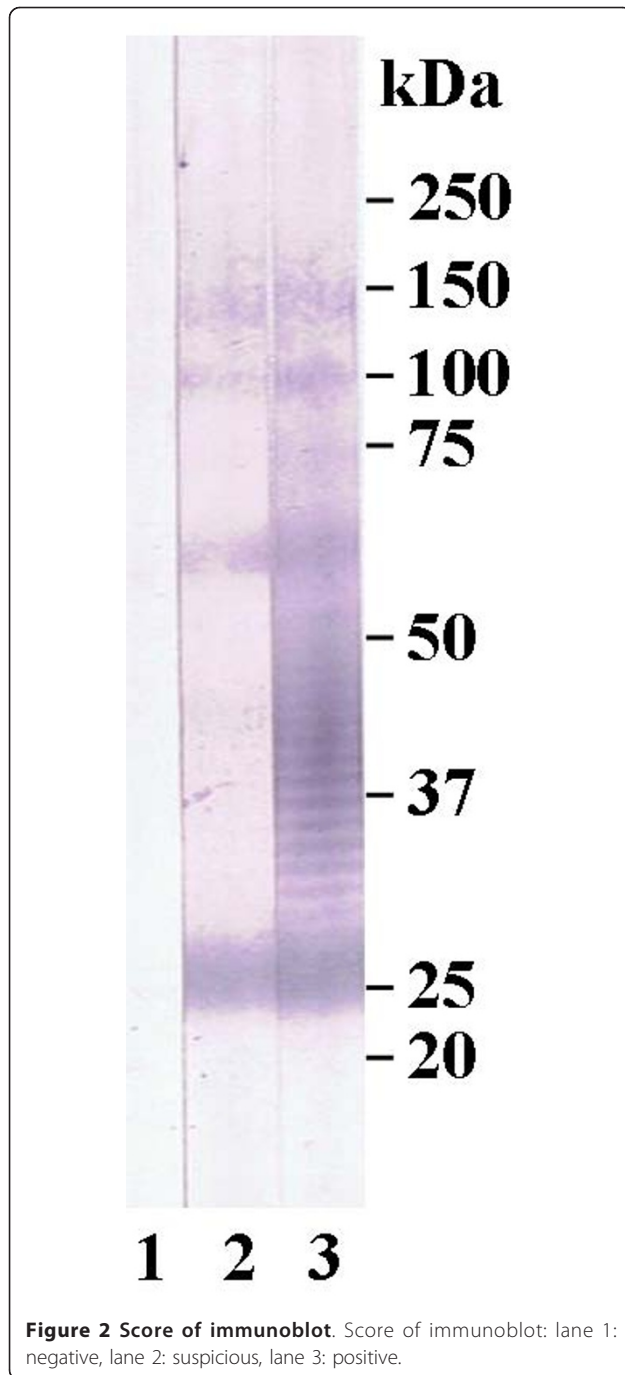
Results

CFT

All 205 samples of group III were positive resulting in a sensitivity of 100%. Seventy of the 1,282 sera of group I showed a false positive or suspicious CFT result resulting in a specificity of 94.5%.

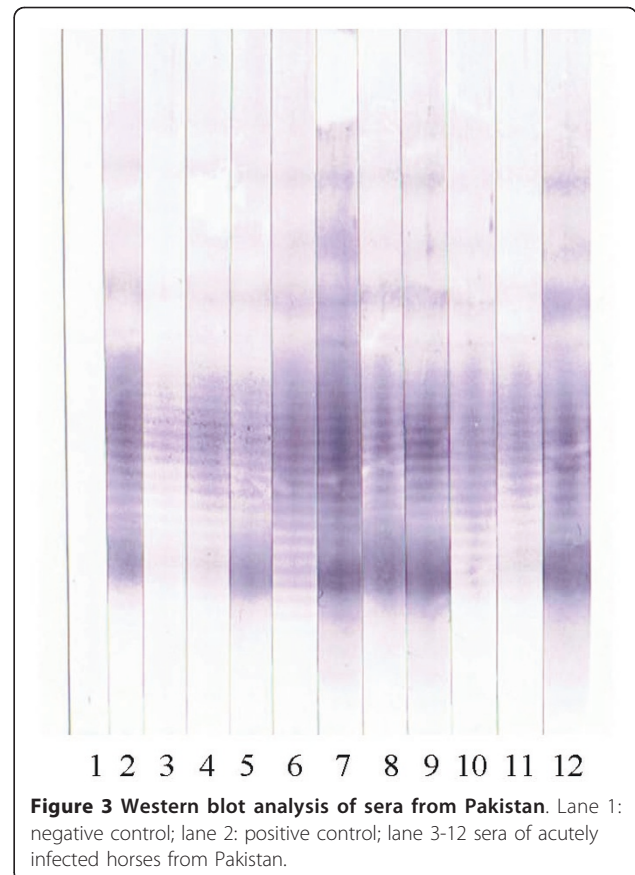
Western blot assay

All 305 samples of group II were tested negative and all of 205 positive samples (group III) showed a clearly



visible LPS banding pattern and were scored positive. A panel of immunoblot strips of samples from Pakistan are shown in Figure 3. On this basis, the diagnostic sensitivity and specificity of the Western blot was calculated to be 100%. CFT false positive samples (n = 70) from group I were also tested negative.

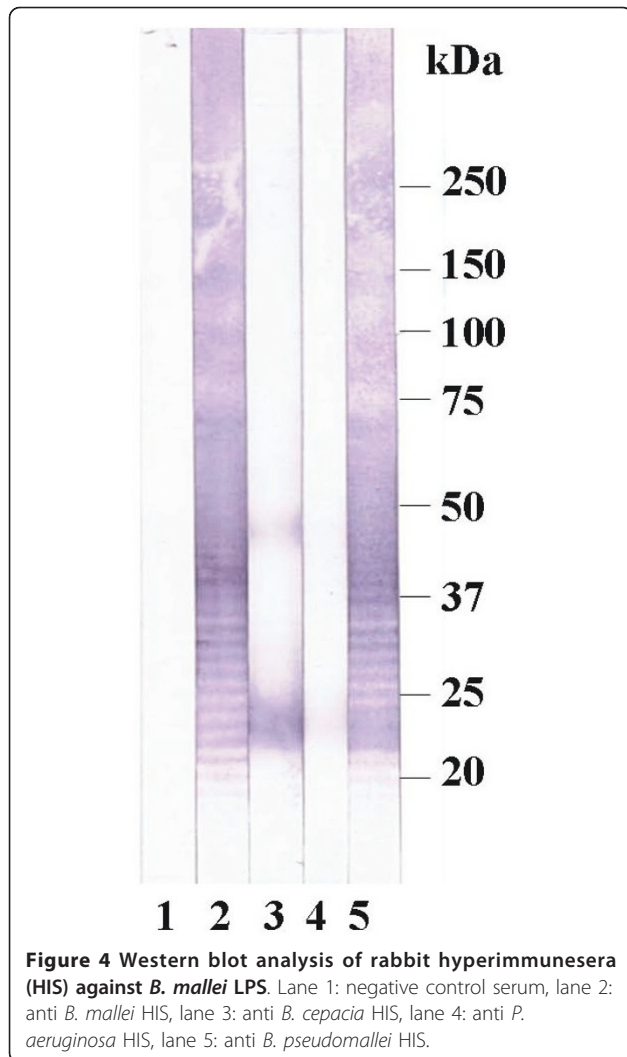
Rabbit hyperimmune sera against *B. cepacia* and *P. aeruginosa* showed no clearly visible LPS banding pattern. Only the anti- *B. pseudomallei* serum cross



reacted with the *B. mallei* antigen in the Western blot assay (Figure 4).

Discussion

Following eradication of glanders from the Western hemisphere, false positive CFT results pose a considerable problem in trade of equids. The need for the development and evaluation of highly specific serological tests for exclusion of false positive results became imperative. One competitive ELISA (cELISA) was described based on mallein as antigen and an in house monoclonal antibody (mAb) [22]. Another cELISA used a commercially available *B. mallei* specific mAb (3D11) and a simple carbohydrate preparation of *B. mallei* as antigen [20]. Both tests show comparable sensitivity (98.9% and 98.6%, respectively) and a specificity of 100%. Due to the sophisticated technique, the high costs for mallein and mAbs and the restricted availability of these mAbs, the cELISAs will not be available for routine laboratories in the future. Other tests still used for diagnosing glanders are Rose Bengal agglutination test, indirect hemagglutination test, counter immunoelectrophoresis and indirect fluorescent antibody test [21,25-29] which have either technical shortcomings or have low specificity and/or sensitivity. New approaches



using microarray technology for the serodiagnosis of glanders and melioidosis are based on polysaccharide antigens [30] but these techniques are expensive and not suited for routine mass testing of sera.

The CFT is used for diagnosis of many diseases worldwide and has a high sensitivity, especially when used for glanders serodiagnosis. This finding was again demonstrated by our study. We conclude therefore that the CFT is still the most valuable screening test currently available. But again, the CFT had a specificity of only 94.5% in a set of horse sera from a glanders free region (Germany). Consequently, a considerable number of CFT false positive tested animals must be expected when horses from glanders free regions are tested by the current OIE standards [6]. The need of the development of a supplementary confirmatory test with a high negative likelihood ratio for international trade was stressed again.

B. mallei LPS-based Western blot assays are cheap, easy to produce and user friendly test systems which have been developed in the past, but could not be evaluated on a significant number of positive and negative horse sera until now [12]. We were able to collect 205 sera of infected animals from geographically different regions and a set of sera from immunized animals (horse and rabbit), enabling us to validate the Western blot assay for its use in the routine serodiagnosis of glanders.

It was proven that the Western blot technique is highly specific and is able to supplement the CFT to avoid false positive diagnoses in horses and mules. The advance of the immunoblot technique is that specific bands can be distinguished from background reactions in contrast to the CFT. The investigation of the hyperimmune sera of rabbits against *B. mallei* LPS showed that *B. cepacia* and *P. aeruginosa* won't interfere with this technique. *B. pseudomallei* antibodies, however, will cause false positive results due to known cross-reacting epitopes of the LPS [31,32]. In international trade, it is also of utmost importance not to transport animals infected with the dangerous zoonotic agent *B. pseudomallei* from endemic areas to areas with ambient environment where *B. pseudomallei* could establish a temporal or permanent reservoir. An identification and destruction of those animals will have an additional zoonosanitary and ecological benefit.

Conclusions

The developed method of LPS-preparation reported here is highly feasible and cheap. The qualitative and quantitative composition of our antigen proved to be suitable for its use in immunoblotting. The developed method is a highly sensitive and specific serological test for glanders diagnosis. It will be of great value in glanders endemic areas in less developed countries.

Abbreviations

CFT: complement fixation test; LPS: lipopolysaccharide; OIE: Office International des Epizooties (World organisation for animal health); Cfu: colony forming units; PBS: phosphate buffered saline; SDS-PAGE: sodium dodecylsulfate polyacrylamide gel electrophoresis; mAb: monoclonal antibody; cELISA: competitive enzyme linked immunosorbent assay; ATCC: American Type Culture Collection; DSM: German Collection of Microorganisms and Cell Cultures

Acknowledgements

We are grateful to C. Lodri and U. Pfeil for excellent technical assistance and to Dr. W. Herbst (Justus-Liebig-University Gießen) for providing collection of horse sera from Germany.

Author details

¹Friedrich Loeffler Institute, Federal Research Institute for Animal Health, Institute of Bacterial Infections and Zoonoses, Naumburger Strasse 96a, 07743 Jena, Germany. ²Bundeswehr Institute of Microbiology, Neuherbergstrasse 11, 80937 Munich, Germany. ³Veterinary Research Centre, PO Box 50 Seeb, Postal Code 121, Sultanate of Oman. ⁴Department of

Clinical Medicine and Surgery, University of Agriculture, Faisalabad 38040, Pakistan. ⁵Laboratório Nacional Agropecuário, Ministério Da Agricultura, Rua Dom Manoel De Medeiros, S/N Dois Irmãos Recife PE Brasil CEP 52171-030, Brazil. ⁶Central Veterinary Research Institute, Dubai PO Box 597, United Arab Emirates.

Authors' contributions

MCE designed the study, coordinated the investigation of all samples by CFT and western blot assay, evaluated and interpreted the data and wrote the manuscript.

HN has the supervision of the research group, contributed to acquisition of funding, co-design of the study and was involved in the drafting of the manuscript.

HCS, HN established the antigen purification procedure and western blot assay.

UW, RW, MS, VLAS, MMAS have been involved in malleinisation and health monitoring of the horses, collection and pre-testing and providing of samples including the interpretation of results.

PM established, standardized and performed the western blot assay.

FM, AR, MD, GS have been involved in the design of the study, providing and supporting laboratory work and discussion of results including drafting the manuscript.

All authors revised the manuscript critically and approved the final version.

Received: 9 August 2010 Accepted: 19 January 2011

Published: 19 January 2011

References

1. Bazargani TT, Tadjbakhs H, Badii A, Zahraei T: **The outbreak of glanders in some racehorses in three states of Iran.** *J Eq Vet Sci* 1996, **16**:232-236.
2. Al-Ani FK, Al-Rawashdeh OF, Ali AH, Hassan FK: **Glanders in horses - clinical, biochemical and serological studies in Iraq.** *Vet Arhiv* 1998, **68**:155-162.
3. Muhammad G, Khan MZ, Athar M: **Clinico-microbiological and therapeutic aspects of glanders in equines.** *J Equine Sci* 1998, **9**:93-96.
4. Arun S, Neubauer H, Gurel A, Ayyidiz G, Kuscus B, Yesidere T, Meyer H, Hermanns W: **Equine glanders in Turkey.** *Vet Rec* 1999, **144**:255-258.
5. Mota RA, Brito MF, Castro FJ, Massa M: **Glanders in horses and mules of the states of Pernambuco and Alagoas, Brazil.** *Pes Vet Bras* 2000, **20**:155-159.
6. Wernery U, Zachariah R, Wernery R, Joseph S, Valsini L: **Ten years of freedom from notifiable equine diseases in the United Arab Emirates.** In *Proceedings of the 15th International Conference of Racing Analysts and Veterinarians 2005; Dubai, United Arab Emirate*. Edited by: Albert PH, Morton T, Wade JF. R 2005:1-4.
7. Odontsetseg N, Mweene AS, Kida H: **Viral and bacterial diseases in livestock in Mongolia.** *Jpn J Vet Res* 2005, **52**:151-162.
8. Malik P, Khurana SK, Singh BK, Dwivedi SK: **Recent outbreak of glanders in India.** *Ind J Animal Science* 2009, **79**:1015-1017.
9. Elschner MC, Klaus CU, Liebler-Tenorio E, Schmoock G, Wohlsein P, Tinschmann O, Lange E, Kaden V, Klopffleisch R, Melzer F, Rassbach A, Neubauer H: **Burkholderia mallei infection in a horse imported from Brazil.** *Equine vet Educ* 2009, **21**:147-150.
10. Naureen A, Saqib M, Muhammad F, Ahmad R, Muhammad G, Asi MN, Hussain MH, Lodhi LA, Khan MS, Thibault FM: **Antimicrobial susceptibility of 41 Burkholderia mallei isolates from spontaneous outbreaks of equine glanders in Punjab, Pakistan.** *J Eq Vet Sci* 2010, **30**:134-141.
11. OIE-WAHID, World Animal Health Information Database Interface of OIE, online version. [http://www.oie.int].
12. Neubauer H, Sprague LD, Zacharia R, Tomaso H, Al Dahouk S, Wernery R, Wernery U, Scholz HC: **Serodiagnosis of Burkholderia mallei infections in horses: state-of-the-art and perspectives.** *J Vet Med B Infect Dis Vet Public Health* 2005, **52**:201-205.
13. OIE: **Glanders.** 2008, Chapter 2.5.11: [http://www.oie.int], online version.
14. Cravitz L, Miller WR: **Immunological studies with Malleomyces mallei and Malleomyces pseudomallei. II. Agglutination and complement fixation tests in man and laboratory animals.** *J Infect Dis* 1950, **86**:52-62.
15. Manninger R: **Rotzkrankheit. Malleus.** In *Spezielle Pathologie und Therapie der Haustiere. Volume 1.* 9 edition. Edited by: Marek J, Manninger R, Mocsy J. Jena: Gustav Fischer; 1945:591-627.
16. Wilson GS, Miles AA: **Glanders and Melioidosis.** In *Topley and Wilson's Principles of Bacteriology and Immunity.* 3 edition. Edited by: Wilson GS, Miles AA. London: Edward Arnold 1946:1413.
17. Misra VC, Arora AK: **Serological cross-reactions between Pseudomonas mallei and some other bacteria.** *Ind J Comp Microbiol Immunol Infect Dis* 1990, **11**:28-31.
18. Turnbull A, Wernery U, Wernery R, Anandh JP, Kinne J: **Survey of six infectious diseases of feral donkeys in United Arab Emirates.** *Eq Vet J* 2002, **14**:33-38.
19. Derbyshire JB: **Eradication of glanders in Canada.** *Can Vet J* 2002, **43**:722-726.
20. Sprague LD, Zachariah R, Neubauer H, Wernery R, Joseph M, Scholz HC, Wernery U: **Prevalence-dependent use of serological tests for diagnosing glanders in horses.** *BMC Vet Res* 2009, **5**:32.
21. Katz J, Chieves LP, Hennager SG, Nicholson J, Fisher TA, Byers PE: **Serodiagnosis of equine piroplasmiasis, dourine, and glanders using an arrayed immunoblotting method.** *J Vet Diagn Invest* 1999, **11**:292-294.
22. Katz J, Dewald R, Nicholson J: **Procedurally similar competitive immunoassay systems for the serodiagnosis of Babesia equi, Babesia caballi, Trypanosoma equiperdum and Burkholderia mallei infection in horses.** *J Vet Diagn Invest* 2000, **12**:46-50.
23. Anuntagool N, Sirisinha S: **Antigenic relatedness between Burkholderia pseudomallei and Burkholderia mallei.** *Microbiol Immunol* 2002, **46**:143-150.
24. Pitt TL, Aucken H, Dance DA: **Homogeneity of lipopolysaccharide antigens in Pseudomonas pseudomallei.** *J Infect* 1992, **25**:139-146.
25. Naureen A, Saqib M, Muhammad G, Hussain MH, Asi MN: **Comparative evaluation of Rose Bengal plate agglutination test, mallein and some conventional serological tests for diagnosis of spontaneous equine glanders.** *J Vet Diagn Invest* 2007, **19**:362-367.
26. Misra VC, Arora AK: **Diagnosis of glanders by counter-immuno-electrophoresis test and its efficacy vis-a-vis other test.** *J Remount Vet Corps* 1988:185.
27. Jana AM, Gupta AK, Pandya G, Verma RD, Rao KM: **Rapid diagnosis of glanders in equines by counter-immuno-electrophoresis.** *Ind Vet J* 1982, **59**:5-9.
28. Ma CL, Fan SM, Wang X, Jiang L, Fang SQ: **Diagnosis of glanders in horses by the indirect fluorescent antibody (IFA) technique.** *Chinese J Vet Sci Technol* 1986, **9**:3-5.
29. Zhang WD, Lu ZB: **Application of an indirect haemagglutination test for the diagnosis of glanders and melioidosis.** *Chinese J Vet Med* 1983, **9**:8-9.
30. Parthasarathy N, DeShazer D, England M, Waag DM: **Polysaccharide microarray technology for the detection of Burkholderia pseudomallei and Burkholderia mallei antibodies.** *Diag Microbiol Infect Dis* 2006, **56**:329-332.
31. Brett PJ, Woods DE: **Structural and immunological characterization of Burkholderia pseudomallei O-polysaccharide-flagellin protein conjugates.** *Infect Immun* 1996, **64**:2824-2828.
32. Burtnick MN, Brett PJ, Woods DE: **Molecular and physical characterization of Burkholderia mallei O antigens.** *J Bacteriol* 2002, **84**:849-852.

doi:10.1186/1746-6148-7-4

Cite this article as: Elschner et al.: Use of a Western blot technique for the serodiagnosis of glanders. BMC Veterinary Research 2011 7:4.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

